

褪黑素调控动物消化道上皮养分吸收代谢机制的研究进展

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摘要: 褪黑素(melatonin, MLT)作为一类主要在松果体内合成的吲哚类激素, 具有抗氧化功能, 同时还能在肠道内合成, 参与消化道的生理功能调控。本文主要从消化道褪黑素来源、代谢规律, 以及其调控脂肪、糖类和氨基酸吸收代谢等方面进行综述, 为褪黑素调控消化道上皮养分吸收代谢的研究提供参考。

关键词: 褪黑素(MLT); 消化道上皮; 养分; 吸收; 代谢

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Research Progress in Mechanism of Regulation of Nutrient Uptake and Metabolism by Melatonin in Animal Digestive Tract Epithelium

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Abstract: Melatonin (MLT), as a class of indole-like hormones mainly synthesized in pinecones, has antioxidant function. It is also synthesized in the intestinal tract and participates in regulating physiological functions of the digestive tract. Herein, the sources and metabolism of melatonin in the digestive tract, and its regulation on the absorption and metabolism of fat, carbohydrate and amino acid were reviewed, which would provide reference for research on melatonin in regulating nutrient absorption and metabolism in the digestive tract epithelium.

Key words: melatonin (MLT); digestive tract epithelium; nutrient; absorption; metabolism

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褪黑素(melatonin, MLT)是一种由 L-色氨酸酶促形成的吲哚类激素, 不仅在松果体内, 而且在无脊椎动物和脊椎动物的胃肠道中产生, 且普遍存在。作为强有力的抗氧化剂, 褪黑素可通过调节前列腺素刺激十二指肠和胰腺分泌碳酸氢根^[1-2], 最终抑制胃蛋白酶和胃酸的分泌, 增加胃黏膜血流。Morais 等^[3]在一项小鼠实验中发现, 低浓度褪黑素可以显著缩短实验组小鼠回肠排除玻璃珠的时间, 增强肠道蠕动。此外, 褪黑素可促进消化道溃疡愈合, 对胃有保护作用, 其不足会影响细胞增殖并损伤肠道黏膜^[4-5]。因此, 研究褪黑素对胃

肠道消化和吸收, 以及胃肠道健康都有着重要意义。本文主要综述了消化道褪黑素的来源与分布、受体表达及其对肠道上皮细胞吸收的调控, 期为褪黑素在调控营养吸收和肠道健康中的应用提供参考。

1 消化道组织中褪黑素的来源与分布

褪黑素最早于 1958 年在牛的松果体中被发现, 1974 年其首次在人的阑尾中被发现^[6]。除松果体外, 胃肠道也是产生褪黑素的重要组织器官。褪黑素在人胃肠道组织中的含量大概是松果体中

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的400倍,主要由位于肠上皮的肠嗜铬细胞合成,且其含量和细胞密度呈正相关,这些褪黑素涉及胃肠道炎症、动力等过程^[7]。相关研究在鹌鹑^[8]和小鼠^[9]的肠道组织中发现了参与合成褪黑素关键酶的基因表达,说明胃肠道组织可能通过自身合成并分泌褪黑素,从而维持胃肠道组织内褪黑素的浓度。

放射性标记2-[¹²⁵I]褪黑素可以在鸭的肠道中使用,用来表征和定位褪黑素的结合位点。基于该项技术有研究发现,在鸭肠的不同区域,2-[¹²⁵I]褪黑素结合位点的密度由低到高依次为:盲肠与食道、结肠、空肠与十二指肠、回肠,这表明褪黑素在鸭肠不同部位分布不均;同时,2-[¹²⁵I]褪黑素在鸭肠中的位点分布没有日节律性^[10]。在哺乳动物的肠道中,褪黑素的分布也存在区域差异性。褪黑素在空肠和回肠中的含量最低,而在直肠和结肠中的含量则相反,这些差异已在人类、兔和小鼠中得到证实^[11]。另外,Bubenik等^[12]测定了不同物种(牛和猪)消化道组织中褪黑素的含量,结果表明:与牛相比,猪的胃和回肠组织中的褪黑素含量要低得多,而猪的盲肠和结肠组织中的褪黑素含量要高得多;牛的黏膜肌层与黏膜层中的褪黑素含量相差不大。

有研究报道,切除大鼠的松果体后,消化系统不同区域的褪黑素水平没有发生变化,而在门静脉部分结扎后,褪黑素水平有减少^[13],表明褪黑素可独立于松果体在消化系统中存在。另有研究显示,与老年小鼠相比,幼年小鼠结肠远端黏膜和回肠的褪黑素含量低126%^[14];大鼠胃肠道组织中的褪黑素含量在出生时达到峰值,在21d时下降至稳定水平^[15],其中空肠、回肠和结肠组织中的褪黑素含量下降更为明显^[16],但在后期胃肠道褪黑素含量增加。这表明,年龄对肠道褪黑素含量有一定的影响。此外,褪黑素的分泌与进食也有一定的关系。Bubenik等^[17]将猪作为研究对象,让其禁食30h后进食,并在进食0h、1h、2h、5h、10h、20h时测定胃肠道组织和血清的褪黑素水平,结果显示,进食后,除直肠褪黑素水平无明显变化以外,各消化道组织和血清中的褪黑素水平均显著升高。

2 消化道组织中褪黑素受体的分布与表达规律

褪黑素的受体在胃肠道组织中也广泛存在^[18],

胃肠系统中的褪黑素受体与位于中枢神经系统中的受体显示出非常相似的特征,具有相似的药理学特异性,因此它们被认为具有相同的性质。褪黑素受体1(melatonin receptor 1, MT1)、MT2和MT3都可在胃肠道表达,其中,作为G蛋白偶联受体家族成员的MT1和MT2,具有7次跨膜结构,在纳摩尔范围内具有高亲和力^[19-20]。有研究发现,褪黑素还可与细胞核中类视黄醇X受体(retinoid X receptor, RXR)或类视黄醇孤儿核受体a(retinoid orphan receptor a, RORa)结合^[21-22],外周血单个核细胞中ROR/RZR基因表达减少伴随着MT1受体mRNA表达的减少,这体现出褪黑素的膜受体和核受体之间存在相互作用。

褪黑素受体的表达在组织和细胞中呈现差异性。刘文举等^[23]采用实时荧光定量PCR和免疫组织化学技术,研究了鸭不同组织中MT1b(melatonin receptor 1b) mRNA和蛋白质的表达分布,发现MT1b mRNA在心脏、肾脏、大脑、卵巢、胸肌、脾脏、肺、肝脏、胰脏中均有表达,且MT1b蛋白也存在于大脑、肺、肝脏、胸肌、肾脏、心脏、胰脏。此外,尽管人的胃肠道血管、黏膜下层、肌间神经丛和肠上皮都有MT1和MT2的表达,但是它们在大肠上皮中的表达量最高,肠嗜铬细胞也存在MT2高表达。褪黑素还可以直接与钙调蛋白、钙网蛋白或微管蛋白等细胞内蛋白质相互作用,这扩展了褪黑素结合和作用的潜在位点^[24]。5-羟色胺(5-hydroxytryptamine, 5-HT)受体也可能是褪黑素的作用位点。Kasimay等^[25]研究表明,先注射5-HT3受体(ramosetrone, 50 g/kg)或胆囊收缩素(CCK2, 1 mg/kg)阻断剂15 min,再添加褪黑素,可消除褪黑素引起的胃排空延迟。这一效应中,褪黑素可能通过阻断烟碱乙酰胆碱受体,或调节细胞膜的Ca²⁺通道发挥作用。

3 胃肠道组织中褪黑素的代谢规律

内源性褪黑素(血浆中游离褪黑素)在血清中的半衰期是30~60 min,外源性褪黑素(静脉注射)的半衰期更短,只有12~48 min^[26]。研究显示,褪黑素静脉注射小鼠后会快速分布到机体的各个器官,其中肝脏和肾脏的含量最高,脂肪和皮肤的含量最低;30 min后,褪黑素在血浆和肝脏的分布量为原来的10%~20%,而在脑肾上腺中的含量为原来的66.7%^[27-28]。

在肠道组织中,褪黑素的前体物质L-色氨

酸, 经色氨酸羟化酶(tryptophan hydroxylase, TPH)催化生成 5-羟色氨酸, 后者经芳香族 L-氨基酸脱羧酶(aromatic L-aminoacid decarboxylase, AA-DC)、5-羟色胺-N-乙酰转移酶(serotonin N-acetyltransferase, SNAT)催化依次生成 5-羟色胺(5-HT)和乙酰 5-羟色胺, 最后乙酰 5-羟色胺在乙酰血清素甲基转移酶(acetyl-serotonin-O-methyltransferase, ASMT)催化下合成褪黑素^[29]。此外, 褪黑素经羟基化代谢生成 6-羟基褪黑素(6-hydroxymelatonin, 6-HMT), 后者的血液浓度比褪黑素高 37 倍, 这一代谢途径由细胞色素 P450 1A1、1A2、1B1 及 2C9 介导^[30-31]。

4 褪黑素对消化道养分吸收代谢的调控

褪黑素作为生理激素, 和其他激素相比, 具有更高的亲脂性, 其发挥作用的方式是多渠道的, 既可以通过结合细胞膜表面的膜受体向细胞内释放第二信号分子, 也可直接进入细胞膜, 与细胞核内的受体结合, 还可经黏膜层与黏膜下层进入到更深层, 然后在肌间神经丛和肌层黏膜发挥作用^[32]。大鼠结肠、胃、十二指肠肌肉层和肌肉黏膜均有褪黑素参与调节胃肠道运动^[33], 但具体的调控机制尚不清楚。褪黑素对胃肠道分泌的调节主要在于影响十二指肠的分泌反应, 首先苯肾上腺素刺激中枢神经系统, 引起肠黏膜释放褪黑素, 随后褪黑素通过旁分泌作用于十二指肠细胞, 进而活化促分泌神经元, 使得碳酸氢盐分泌量升高; 褪黑素也可以通过增加十二指肠细胞内钙离子的浓度, 激发碳酸氢离子与氯离子的交换^[34]。鸟类中的研究也发现, 褪黑素能够直接调控鸟类胃肠的功能: 肠道褪黑素释放进入全身循环后, 可使胃腺黏膜和上皮钠的吸收减少, 从而抑制空肠上皮的增殖^[35]。

4.1 褪黑素对脂肪代谢的调控

在营养分配和代谢方面, 褪黑素可以降低动物采食量, 影响能量代谢和动物生长。研究证明, 褪黑素既可以通过受体介导机制降低细胞内甘油三酯的合成和沉积, 又可以降低机体胆固醇的合成, 从而减少血脂的含量^[36]。闫灵敏等^[37-38]研究了褪黑素对大鼠脂肪代谢相关酶表达量的影响, 结果表明, 褪黑素处理后, 脂肪分解有关的酶肉毒碱棕榈酰转移酶 1 (carnitine palmitoyltransferase 1, CPT1)以及转录因子 ALDH3A2 (aldehyde dehydrogenase 3 family member A2)的表达量升高, 而脂肪

合成相关的酶长链脂酰辅酶 A 合成酶 3 (long-chain acyl-CoA synthetase 3, ACSL3)、脂肪酸合成酶(fatty acid synthase, FAS)的 mRNA 表达量降低。不仅如此, 近年来, 越来越多的研究发现, 褪黑素能够调控肠道微生物的脂质代谢过程。Yin 等^[39]报道, 褪黑素可通过影响饲喂高脂食物小鼠的肠道微生物区系, 促进短链脂肪酸的代谢。Xu 等^[40]也发现, 小鼠体内的褪黑素可以通过改变肠胃微生物区系结构抑制肥胖。

相关研究报道, 褪黑素注射液可显著增加蟹肝胰腺中脂肪酶的活性, 加速脂质代谢, 促进肠道消化^[41]。此外, 有研究证明, 褪黑素通过抑制 Notch 信号通路, 在调节鸡肠黏膜结构的完整性以及消化吸收功能中起着关键作用^[42]。综上可知, 褪黑素对脂肪代谢有重要意义。

4.2 褪黑素对糖吸收代谢的调控

褪黑素被认为是葡萄糖代谢的重要调节剂。正常情况下, 下丘脑下部前端的视交叉上核调节褪黑素的分泌模式, 使其呈现昼伏夜出的节律性变化, 而胰岛素呈现出和褪黑素相反的昼高夜低的脉冲式分泌模式^[43]。胰岛素通过调节血糖水平, 在机体的糖代谢中起着决定性的作用。视网膜上的光传感器受到光线刺激, 控制位于下丘脑前部的视交叉上核的昼夜节律。白天, 去甲肾上腺素发挥效应, 降低颈上神经节到松果体的传入冲动, 阻碍肾上腺素受体与兴奋性 G 蛋白偶联, 降低腺苷酸环化酶(adenylyl cyclase, AC)和环磷酸腺苷(cyclic adenosine monophosphate, cAMP)的水平, 抑制 cAMP 依赖蛋白激酶 A (protein kinase A, PKA)激活, 最终激活泛素蛋白酶体系统, 加速 5-羟色胺-N-乙酰转移酶(SNAT)的降解, 使得褪黑素的合成减少, 同时胰腺 β 细胞分泌胰岛素增加^[44], 激素水平的变化刺激组织细胞对血液中葡萄糖的吸收, 并使机体将吸收的葡萄糖合成肌糖原或转化为脂肪, 从而降低血糖水平。有研究显示, 褪黑素可以显著促进葡萄糖转运蛋白 5 (glucose transporter, GLUT5)的 mRNA 在十二指肠的表达^[45]。胰腺 β 细胞中表达的 G 蛋白偶联受体为褪黑素受体 MT1 及 MT2, 它们与抑制性 G 蛋白(Gi)相偶联来介导对胰腺 β 细胞功能和胰岛素分泌发挥作用的 3 个平行信号通路^[46]。MT1 与 G 蛋白(Gq)偶联即可活化磷脂酶 C (phospholipase C, PLC), 使得位于细胞膜上的 4,5-二磷脂酰肌醇(phosphatidylinositol-4,5-bisphosphate, PIP2)分解成三磷酸肌醇(inositol

trisphosphate, IP3)与二酰甘油(diacylglycerol, D-AG)^[47], IP3 进一步促进内质网释放出 Ca^{2+} 作用于致密核心大囊泡(large dense core vesicle, LDCV), 引起胰岛素分泌水平升高^[48]。MT2 与 G_q 偶联的最终效果与 MT1 相反, 其通过降低环磷酸鸟苷(cyclic guanosine monophosphate, cGMP)水平导致胰岛素分泌减少。此外, MT1 和 MT2 可与 G_i 分别偶联, 偶联的结果是胞内 cAMP 水平降低, PKA 活性被抑制, 同时胰高血糖素样肽-1 (glucagon-like peptide-1, GLP-1)和胃抑制肽(gastric inhibitory peptide, GIP)^[49]的分泌水平降低, 引起胰岛素分泌减少^[50-51]。

关于褪黑素对小鸡肠道影响的研究表明, 褪黑素处理不仅增加了十二指肠的麦芽糖酶和蔗糖酶活性, 而且还提高了空肠的蔗糖酶和乳糖酶活性^[52], 提示褪黑素可以增强小鸡肠道消化吸收功能。需要指出的是, 目前有关褪黑素对反刍动物消化和吸收功能的研究尚未见报道, 值得进一步关注。

4.3 褪黑素对蛋白质和氨基酸吸收代谢的调控

褪黑素影响蛋白质消化以及转运蛋白的表达。有研究使用 qRT-PCR 方法检测褪黑素处理后鸡肠道中营养转运蛋白基因的表达, 结果显示, 褪黑素不仅可显著促进 GLUT5 mRNA 在十二指肠的表达, 而且还促进了中性氨基酸转运体 LAT (L-type amino acid transporter) mRNA 在十二指肠的表达, 另外 LAT 和阴离子氨基酸转运蛋白 EA-AT3 (excitatory amino acid transporter 3)的 mRNA 在雏鸡空肠中的表达也显著增加^[53]。这表明, 褪黑素能够促进十二指肠中性氨基酸的吸收。但唐春祥等^[54]通过研究褪黑素对生长猪粗蛋白质消化率的影响发现, 高褪黑素组的粗蛋白质表观消化率差异显著并呈现降低的趋势, 低褪黑素组粗蛋白质表观消化率提高。Konturek 等^[55]的研究结果提示, 褪黑素对蛋白质的消化调控主要是通过减少胃酸分泌量来实现的。

褪黑素促进氨基酸的吸收。在一定程度上, 动物吸收氨基酸的量可以由血浆游离氨基酸直接体现, 且两者呈现正相关, 即饲料中的氨基酸在动物小肠被吸收后, 血浆游离氨基酸含量会上升^[56-57]。Ma 等^[58]按 4 g/d 的剂量在辽宁绒山羊饲料中添加过瘤胃色氨酸(L-色氨酸含量 33%), 结果显示血浆色氨酸含量呈现显著升高的趋势。Kollmann 等^[59]按 500 g/d 的剂量在奶牛饲料中添加过瘤胃色氨

酸(L-色氨酸含量 25%), 结果显示白天与夜晚的血浆色氨酸含量都呈现显著升高的趋势, 但夜间增加更为显著, 具有明显的昼夜节律性。另外, 杨春合等^[60-61]通过给母羔羊埋植褪黑素(2 mg/kg)发现, 母羊血清中褪黑素含量显著增加, 而催乳素含量则显著降低, 母羊的产奶量也显著降低。

综上所述, 褪黑素可能通过影响胃酸分泌降低蛋白质的消化, 通过包被或其他保护技术可在一定程度上利用褪黑素调控产奶量。

5 结语

褪黑素作为重要的生理激素, 可调控消化道上皮的糖脂代谢。虽然褪黑素可通过影响胰脂肪酶、肠道菌群来调控脂肪的吸收与代谢, 但褪黑素对脂肪组织本身是否产生影响及其调节脂肪代谢的具体通路还不清楚; 同样, 虽然有研究表明褪黑素可通过调节胰岛素分泌以及麦芽糖酶、蔗糖酶和乳糖酶的活性来调节糖的吸收和代谢, 但需要指出的是, 目前有关褪黑素对反刍动物糖代谢的研究尚未见报道, 值得进一步关注。当前, 针对褪黑素的研究主要集中在基于大鼠模型的抗氧化等生理功能探究; 对于消化道上皮的吸收作用只停留在生物钟这一层面, 利用褪黑素研究消化道上皮吸收的报道比较少。总的来讲, 褪黑素调控肠上皮细胞和组织受体的信号通路, 促进糖类、脂肪、氨基酸的吸收代谢机制有待进一步阐明, 在消化道内如何靶向调控褪黑素受体发挥生物学效应将是该领域研究的重点。

参考文献(References):

- [1] CARRILLO-VICO A, GUERRERO J M, LARDONE P J, *et al.* A review of the multiple actions of melatonin on the immune system[J]. *Endocrine*, 2005, 27(2): 189-200.
- [2] JAWOREK J, BRZOZOWSKI T, KONTUREK S J. Melatonin as an organoprotector in the stomach and the pancreas[J]. *Journal of Pineal Research*, 2005, 38(2): 73-83.
- [3] MORAIS T C, ARRUDA B R, DE SOUSA MAGALHAES H, *et al.* Mangiferin ameliorates the intestinal inflammatory response and the impaired gastrointestinal motility in mouse model of postoperative ileus[J]. *Naunyn Schmiedeberg's Archives of Pharmacology*, 2015, 388(5): 531-538.
- [4] BRZOZOWSKA I, STRZALKA M, DROZDOWICZ D, *et al.* Mechanisms of esophageal protection, gastroprotection and ulcer healing by melatonin. Implications for the therapeutic use of melatonin in gastroesophageal reflux disease (GERD) and peptic ulcer disease[J]. *Current Pharmaceutical Design*, 2014, 20(30): 4807-4815.
- [5] KONTUREK S J, KONTUREK P C, BRZOZOWSKI T, *et al.* Role of melatonin in upper gastrointestinal tract[J]. *Journal of Physiology and Pharmacology*, 2007, 58(Suppl. 6): 23-52.
- [6] BUBENIK G A. Thirty-four years since the discovery of gastrointestinal melatonin[J]. *Journal of Physiology and Pharmacology*, 2008, 59(Suppl. 2): 33-51.

- [7] FERLAZZO N, ANDOLINA G, CANNATA A, *et al.* Is melatonin the cornucopia of the 21st century[J]. *Antioxidants*, 2020, 9(11): 1088.
- [8] HONG G X, PANG S F. N-Acetyltransferase activity in the quail (*Coturnix coturnix jap*) duodenum[J]. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 1995, 112(2): 251–255.
- [9] STEFULJ J, HÖRNER M, GHOSH M, *et al.* Gene expression of the key enzymes of melatonin synthesis in extrapineal tissues of the rat[J]. *Journal of Pineal Research*, 2001, 30(4): 243–247.
- [10] LEE P P, SHIU S Y, CHOW P H, *et al.* Regional and diurnal studies of melatonin and melatonin binding sites in the duck gastrointestinal tract[J]. *Biomedical Signal Processing and Control*, 1995, 4(4): 212–224.
- [11] POON A M, CHOW P H, MAK A S, *et al.* Autoradiographic localization of 2^{125} Ijodometatonin binding sites in the gastrointestinal tract of mammals including humans and birds[J]. *Journal of Pineal Research*, 1997, 23(1): 5–14.
- [12] BUBENIK G A, HACKER R R, BROWN G M, *et al.* Melatonin concentrations in the luminal fluid, mucosa, and muscularis of the bovine and porcine gastrointestinal tract[J]. *Journal of Pineal Research*, 1999, 26(1): 56–63.
- [13] BUBENIK G A, BROWN G M. Pinelectomy reduces melatonin levels in the serum but not in the gastrointestinal tract of rats[J]. *Biological Signals*, 1997, 6(1): 40–44.
- [14] BERTRAND P P, BERRAN R L, CAMELLO P J, *et al.* Simultaneous measurement of serotonin and melatonin from the intestine of old mice: the effects of daily melatonin supplementation[J]. *Journal of Pineal Research*, 2010, 49(1): 23–34.
- [15] BUBENIK G A. Localization of melatonin in the digestive tract of the rat[J]. *Hormone Research*, 1980, 12(6): 313–323.
- [16] BUBENIK G A, PANG S F. The role of serotonin and melatonin in gastrointestinal physiology: ontogeny, regulation of food intake, and mutual serotonin-melatonin feedback[J]. *Journal of Pineal Research*, 1994, 16(2): 91–99.
- [17] BUBENIK G A, PANG S F, HACKER R R, *et al.* Melatonin concentrations in serum and tissues of porcine gastrointestinal tract and their relationship to the intake and passage of food[J]. *Journal of Pineal Research*, 1996, 21(4): 251–256.
- [18] BUBENIK G A. Localization, physiological significance and possible clinical implication of gastrointestinal melatonin[J]. *Biological Signals and Receptors*, 2001, 10(6): 350–366.
- [19] DUBOCOVICH M L, MARKOWSHA M. Functional MT1 and MT2, melatonin receptors in mammals[J]. *Endocrine*, 2005, 27(2): 101–110.
- [20] CHEN C Q, FICHNA J, BASHASHATI M, *et al.* Distribution, function and physiological role of melatonin in the lower gut[J]. *World Journal of Gastroenterology*, 2011, 17(34): 3888–3898.
- [21] CARLBERG C. Gene regulat on by melatonin[J]. *Annals of the New York Academy of Sciences*, 2000, 917(1): 387–396.
- [22] HIROSE T, SMITH R J, JETTEN A M. ROR gamma: the third member of ROR/RZR orphan receptor subfamily that is highly expressed in skeletal muscle[J]. *Biochemical and Biophysical Research Communications*, 1994, 205(3): 1976–1983.
- [23] 刘文举, 王淑娟, 刘晓娟, 等. 褪黑素受体 *Mel1b* 基因 mRNA 和蛋白在鸭不同组织中的表达与分布[J]. *浙江农业学报*(LIU Wen-ju, WANG Shu-juan, LIU Xiao-li, *et al.* Expression and distribution of *Mel1b* mRNA and protein in various tissues of duck[J]. *Acta Agriculturae Zhejiangensis*), 2018, 30(5): 711–716.
- [24] SÖDERQUIST F, HELLSTRÖM P M, CUNNINGHAM J L. Human gastroenteropancreatic expression of melatonin and its receptors MT1 and MT2[J]. *PLoS One*, 2015, 10(3): e0120195.
- [25] KASIMAY O, CAKIR B, DEVSEREN E, *et al.* Exogenous melatonin delays gastric emptying rate in rats: role of CCK2 and 5-HT3 receptors[J]. *Journal of Physiology and Pharmacology*, 2005, 56(4): 543–553.
- [26] KENTISH S J, VINCENT A D, KENNAWAY D J, *et al.* High-fat diet-induced obesity ablates gastric vagal afferent circadian rhythms[J]. *The Journal of Neuroscience*, 2016, 36(11): 3199–3207.
- [27] STEBELOVÁ K, ANTTILA K, MÄNTTÄRI S, *et al.* Immunohistochemical definition of MT2 receptors and melatonin in the gastrointestinal tissues of rat[J]. *Acta Histochemica*, 2010, 112(1): 26–33.
- [28] 周汾, 李肇端, 余剑波, 等. 褪黑素在围手术期的应用[J]. *临床麻醉学杂志*(ZHOU Fen, LI Zhao-duan, YU Jian-bo, *et al.* Application of melatonin during perioperative period[J]. *Journal of Clinical Anesthesiology*), 2015, 31(3): 302–304.
- [29] TAN D X, MANCHESTER L C, ESTEBAN-ZUBERO E, *et al.* Melatonin as a potent and inducible endogenous antioxidant: synthesis and metabolism[J]. *Molecules*, 2015, 20(10): 18886–18906.
- [30] 刘哲宇, 孙铮. 褪黑素代谢模式的研究进展 [J]. *生命科学*(LIU Zhe-yu, SUN Zheng. The research progress of melatonin metabolic patterns[J]. *Chinese Bulletin of Life Sciences*), 2017, 29(2): 209–214.
- [31] LI C Y, LI G M, TAN D X, *et al.* A novel enzyme-dependent melatonin metabolite in humans[J]. *Journal of Pineal Research*, 2013, 54(1): 100–106.
- [32] 梅花, 丽春. 褪黑素在动物中的作用研究进展[J]. *饲料工业*(MEI Hua, LI Chun. Research progress of the effort of melatonin in animal[J]. *Feed Industry*), 2017, 38(1): 62–64.
- [33] JAN J E, REITER R J, WONG P K H, *et al.* Melatonin has membrane receptor-independent hypnotic action on neurons: an hypothesis[J]. *Journal of Pineal Research*, 2011, 50(3): 233–240.
- [34] 谭碧娥, 任文凯, 印遇龙. 胃肠道褪黑素分泌及其生理功能[J]. *动物营养学报*(TAN Bi-e, REN Wen-kai, YIN Yu-long. Secretion and physiological functions of melatonin in the gastrointestinal tract[J]. *Chinese Journal of Animal Nutrition*), 2018, 30(4): 1207–1216.
- [35] SJÖBLOM M. The duodenal mucosal bicarbonate secretion[J]. *Upsala Journal of Medical Sciences*, 2005, 110(2): 115–150.
- [36] DE FARIAS T D S M, CRUZ M M, DE SA R C D C, *et al.* Melatonin supplementation decreases hypertrophic obesity and inflammation induced by high-fat diet in mice[J]. *Frontiers in Endocrinology*, 2019, 10: 750.
- [37] JIN C J, ENGSTLER A J, SELLMANN C, *et al.* Sodium butyrate protects mice from the development of the early signs of non-alcoholic fatty liver disease: role of melatonin and lipid peroxidation[J]. *British Journal of Nutrition*, 2016, 116(10): 1682–1693.
- [38] 闫灵敏, 杨改青, 王林枫, 等. 褪黑素调节雄性大鼠脂肪代谢的研究[J]. *畜牧兽医学报*(YAN Ling-min, YANG Gai-qing, WANG Lin-feng, *et al.* The study of melatonin regulating lipid metabolism in male rats[J]. *Acta Veterinaria et Zootechnica Sinica*), 2017, 48(9): 1694–1704.
- [39] YIN J, LI Y Y, HAN H, *et al.* Melatonin reprogramming of gut microbiota improves lipid dysmetabolism in high-fat diet-fed mice[J]. *Journal of Pineal Research*, 2018, 65(4): e12524.
- [40] XU P F, WANG J L, HONG F, *et al.* Melatonin prevents obesity through modulation of gut microbiota in mice[J]. *Journal of Pineal Research*, 2017, 62(4): e12399.
- [41] 徐敏杰. L-色氨酸和褪黑激素对中华绒螯蟹血糖代谢及免疫性能的影响[D]. 上海: 海洋大学(XU Min-jie. Analysis of the Effect of L-Tryptophan and Melatonin on the Hemolymph Glucose Metabolism and Immune Performance in the *Eriocheir sinensis*[D]. Shanghai: Shanghai Ocean University), 2018.
- [42] WANG Y H, KUANG Z, YU X F, *et al.* The intestinal microbiota regulates body composition through NFIL 3 and the circadian clock[J]. *Science*, 2017, 357(6354): 912–916.
- [43] COSTES S, BOSS M, THOMAS A P, *et al.* Activation of melatonin signaling promotes β -cell survival and function[J]. *Molecular Endocrinology*, 2015, 29(5): 682–692.
- [44] AHN J H, PARK J H, KIM I H, *et al.* Comparison of arylalkylamine N-acetyltransferase and melatonin receptor type 1B immunoreactivity between young adult and aged canine spinal cord[J]. *Journal of Veterinary Science*, 2014, 15(3): 335–342.
- [45] KALSBECK A, LA FLEUR S, FLIERS E. Circadian control of glucose metabolism[J]. *Molecular Metabolism*, 2014, 3(4): 372–383.
- [46] 汪娅, 吴红艳. 褪黑素及其受体与糖尿病关系的研究进展[J]. *重庆医学*(WANG Ya, WU Hong-yan. Research progress on the relationship between melatonin and its receptor and diabetes mellitus[J]. *Chongqing Medicine*), 2012, 41(12): 1228–1231.
- [47] NISHIYAMA K, HIRAI K. The melatonin agonist ramelteon induces duration-dependent clock gene expression through cAMP signaling in pancreatic INS-1 β -cells[J]. *PLoS One*, 2014, 9(7): e102073.
- [48] CHOI T Y, KWON J E, DURRANCE E S, *et al.* Melatonin inhibits voltage sensitive Ca^{2+} channel-mediated neurotransmitter release[J]. *Brain Research*, 2014, 1557: 34–42.
- [49] SEGHERI M, REBELOS E, GASTALDELLI A, *et al.* Direct effect of GLP-1 infusion on endogenous glucose production in humans[J]. *Diabetologia*, 2013, 56(1): 156–161.

- [9] 王红刚, 陈银华, 于晓惠, 等. 海南省番茄晚疫病病菌对甲霜灵的敏感性及其有效药剂筛选[J]. 云南农业大学学报(自然科学)(WANG Hong-gang, CHEN Yin-hua, YU Xiao-hui, et al. Sensitivity to metalaxyl and screening of the effective fungicides of *Phytophthora infestans* in Hainan Province[J]. Journal of Yunnan Agricultural University (Natural Science), 2020, 35(6): 957-962, 982.
- [10] 穆凯热姆·阿卜来提, 来娜娜, 王晓东. 放线菌生物防治棉花黄萎病研究进展[J]. 新疆农垦科技(MUKAIJEM Ablati, LAI Na-na, WANG Xiao-dong. Research progress on biological control of cotton verticillium wilt by actinomycetes[J]. Xinjiang Farm Research of Science and Technology), 2016, 39(5): 30-33.
- [11] ABBASI S, SAFAIE N, SADEGHI A, et al. *Streptomyces* strains induce resistance to *Fusarium oxysporum* f. sp. *lycopersici* race 3 in tomato through different molecular mechanisms[J]. Frontiers in Microbiology, 2019, 10: 1505.
- [12] BÉRDY J. Thoughts and facts about antibiotics: where we are now and where we are heading[J]. The Journal of Antibiotics, 2012, 65(8): 385-395.
- [13] NGUYEN H T T, PARK A R, HWANG I M, et al. Identification and delineation of action mechanism of antifungal agents: reveromycin E and its new derivative isolated from *Streptomyces* sp. JCK-6141[J]. Postharvest Biology and Technology, 2021, 182: 111700.
- [14] BOUKAEW S, PLUBRUKAM A, PRASERTSAN P. Effect of volatile substances from *Streptomyces philanthi* RM-1-138 on growth of *Rhizoctonia solani* on rice leaf[J]. BioControl, 2013, 58(4): 471-482.
- [15] CHEN J, XUE Q H, MCERLEAN C S P, et al. Biocontrol potential of the antagonistic microorganism *Streptomyces enissocaealis* against *Orobanche cumand*[J]. BioControl, 2016, 61(6): 781-791.
- [16] ANDRADE-HOYOS P, SILVA-ROJAS H V, ROMERO-ARENAS O. Endophytic *Trichoderma* species isolated from *Persea americana* and *Cinnamomum verum* roots reduce symptoms caused by *Phytophthora cinnamomi* in avocado[J]. Plants, 2020, 9(9): 1220.
- [17] DA SILVA L J, TAKETANI R G, DE MELO I S, et al. *Streptomyces araujoniae* sp. nov.: an actinomycete isolated from a potato tubercle[J]. Antonie Van Leeuwenhoek, 2013, 103(6): 1235-1244.
- [18] SILVA L J, CREVELIN E J, SOUZA W R, et al. *Streptomyces araujoniae* produces a multiantibiotic complex with ionophoric properties to control *Botrytis cinerea*[J]. Phytopathology, 2014, 104(12): 1298-1305.
- [19] LIU L M, ZHAO Y, LU J Y, et al. Identification and evaluation of UV-tolerant *Streptomyces araujoniae* strain isolated from Tibet Plateau soil as biocontrol agent against rice blast[J]. Rice Science, 2020, 27(5): 359-362.
- [20] 卢彩鸽, 董红平, 张殿朋, 等. 解淀粉芽胞杆菌 MH71 摇瓶发酵培养基及发酵条件优化[J]. 中国生物防治学报(LU Cai-ge, DONG Hong-ping, ZHANG Dian-peng, et al. Optimization of fermentation medium components and cultural conditions for *Bacillus amyloliquefaciens* MH71 in flask[J]. Chinese Journal of Biological Control), 2015, 31(3): 369-377.
- [21] 刘一贤, 施玉萍, 戴利铭, 等. 橡胶褐根病拮抗放线菌 17-7 的筛选、鉴定及发酵条件优化[J]. 微生物学通报(LIU Yi-xian, SHI Yu-ping, DAI Li-ming, et al. Screening, identification and fermentation optimization of an antimicrobial actinomycete strain 17-7 to *Phellinus noxius*[J]. Microbiology China), 2020, 47(1): 118-129.
- [22] 郑东光, 周蕊, 李俊州, 等. 疮痂链霉菌许昌亚种 SCY114 发酵条件的研究[J]. 河南农业大学学报(ZHENG Dong-guang, ZHOU Rui, LI Jun-zhou, et al. Optimization of medium and fermentation conditions for *Streptomyces scabiei* subsp. *xuchangensis* SCY114[J]. Journal of Henan Agricultural University), 2013, 47(1): 55-60.
- [23] 张红. 白黄链霉菌 TD-1 抗菌活性物质发酵条件优化及分离纯化[D]. 天津: 天津科技大学(ZHANG Hong. Optimization of Fermentation Conditions and Purification of Antifungal Substances from *Streptomyces alboflavus* TD-1[D]. Tianjin: Tianjin University of Science and Technology), 2013.
- [24] 刘晓飞, 侯艳, 马京求, 等. 降解玉米芯木质纤维素放线菌的筛选与发酵条件优化[J]. 农业机械学报(LIU Xiao-fei, HOU Yan, MA Jing-qiu, et al. Efficient degradation and optimization of fermentation conditions of actinomycetes from corn cob[J]. Transactions of the Chinese Society for Agricultural Machinery), 2020, 51(11): 329-337.
- [25] 杨祁云, 王成, 冼海辉, 等. 海洋放线菌株 H74-18 抗菌活性及其发酵条件优化[J]. 中国海洋药物(YANG Qi-yun, WANG Cheng, XIAN Hai-hui, et al. Antimicrobial activity and fermentation technology of marine actinomycete strain H74-18[J]. Chinese Journal of Marine Drugs), 2020, 39(5): 52-58.
- [26] 周丽娜, 王莉莉, 张永娜, 等. 2 株放线菌的抗菌活性及分类学地位[J]. 中国农学通报(ZHOU Li-na, WANG Li-li, ZHANG Yong-na, et al. Antifungal activity and taxonomic status of two actinomycetes[J]. Chinese Agricultural Science Bulletin), 2015, 31(11): 182-189.
- [27] 朱天辉, 张丽娜, 李姝江, 等. 绛红褐链霉菌 SP3 菌株及其在红豆杉土传根病和红豆杉促生上的应用(ZHU Tian-hui, ZHANG Li-na, LI Shu-jiang, et al. Strain of *Streptomyces purpurea* SP3 and its application in yew soil-borne root disease and growth promotion of yew): CN201210367119.3[P]. 2013-12-18.

(上接第 228 页)

- [50] OTTUM M S, MISTRY A M. Advanced glycation end-products: modifiable environmental factors profoundly mediate insulin resistance[J]. Journal of Clinical Biochemistry and Nutrition, 2015, 57(1): 1-12.
- [51] KUMAR YADAV S, HALDAR C, KUMAR SINGH S, et al. Melatonin regulates splenocytes proliferation via IP3-dependent intracellular Ca²⁺ release in seasonally breeding bird, *Perdica asiatic*[J]. Journal of Receptors and Signal Transduction, 2014, 34(4): 233-240.
- [52] RAMRACHEYA R D, MULLER D S, SQUIRES P E, et al. Function and expression of melatonin receptors on human pancreatic islets[J]. Journal of Pineal Research, 2008, 44(3): 273-279.
- [53] KONTUREK P K, BRZOZOWSKI T, KONTUREK S J, et al. Role of epidermal growth factor, prostaglandin, and sulfhydryls in stress-induced gastric lesions[J]. Gastroenterology, 1990, 99(6): 1607-1615.
- [54] 唐春祥. 褪黑激素主动免疫对生长猪生产性能和胴体品质的影响[D]. 雅安: 四川农业大学(TANG Chun-xiang. Effects of Active Immunization Against Melatonin on the Growth Performance and Carcass Quality of Growing Pigs[D]. Ya'an: Sichuan Agricultural University), 2005.
- [55] KONTUREK P C, KONTUREK S J, MAJKA J, et al. Melatonin affords protection against gastric lesions induced by ischemia-reperfusion possibly due to its antioxidant and mucosal microcirculatory effects[J]. European Journal of Pharmacology, 1997, 322(1): 73-77.
- [56] BRZOZOWSKI T, KONTUREK P, SLIWOWSKI Z, et al. Adaptive cytoprotection by ammonia and urea-urease system in the rat gastric mucosa[J]. Journal of Physiology and Pharmacology, 1995, 46(4): 471-488.
- [57] GIBB D J, KLOPFENSTEIN T J, BRITTON R A, et al. Plasma amino acid response to graded levels of escape protein[J]. Journal of Animal Science, 1992, 70(9): 2885-2892.
- [58] MA H, CHENG J B, ZHU X P, et al. Effects of rumen-protected tryptophan on performance, nutrient utilization and plasma tryptophan in cashmere goats[J]. African Journal of Biotechnology, 2011, 10(30): 5806-5811.
- [59] KOLLMANN M T, LOCHER M, HIRCHE F, et al. Effects of tryptophan supplementation on plasma tryptophan and related hormone levels in heifers and dairy cows[J]. Domestic Animal Endocrinology, 2008, 34(1): 14-24.
- [60] 杨春合, 张微, 付霞杰, 等. 褪黑素埋植对内蒙古绒山羊母羊产奶性能、羔羊毛囊发育和产绒性能的影响[C]//2018 年全国养羊生产与学术研讨会论文集. 蚌埠: 中国畜牧兽医学学会养羊学分会(YANG Chun-he, ZHANG Wei, FU Xia-jie, et al. Effects of melatonin implantation on milking performance, hair follicle development and cashmere performance of Inner Mongolia cashmere goat ewes[C]//Proceedings of the 2018 National Sheep Production and Academic Symposium. Bengbu: Sheep Raising Branch of Chinese Association of Animal Science and Veterinary Medicine), 2018: 196.
- [61] 陈俊宏, 赵芳, 魏凯敏, 等. 添喂色氨酸、过瘤胃色氨酸对奶牛泌乳性能、血浆指标和乳中褪黑素含量的影响[J]. 动物营养学报(CHEN Jun-hong, ZHAO Fang, WEI Kai-min, et al. Effects of tryptophan and rumen-protected tryptophan supplementations on lactation performance, plasma indexes and milk melatonin content of dairy cows[J]. Chinese Journal of Animal Nutrition), 2017, 29(11): 3921-3931.